

Application of the Arbuzov Reaction for the Synthesis of Phosphonate Analogues of *Myo*-inositol 1,2-bis- and 1,2,6-trisphosphates and Methyl α -D-mannopyranoside 2,3,4-trisphosphate

Grzegorz Salamończyk^a, Nicola Rehnberg^b, Bożena Krawiecka^a and Jan Michalski^{a*}

^a Centre of Molecular and Macromolecular Studies, Polish Academy of Sciences, Sienkiewicza 112, 90-363 Łódź, Poland

^b Perstorp Pharma, S-284 80 Perstorp, Sweden

Abstract: The reaction of perbenzylated (\pm)-*myo*-inositol 1,2-bis- and 1,2,6-tris-phosphites with benzyl bromoacetate, followed by catalytic (Pd/C) hydrogenolysis affords (\pm)-*myo*-inositol 1,2-bis- and 1,2,6-tris(carboxymethylphosphonate). The same procedure is used for the synthesis methyl α -D-mannopyranoside 2,3,4-tris(carboxymethylphosphonate).
© 1997, Published by Elsevier Science Ltd. All rights reserved.

Recognition of Ins(1,4,5)P₃ as a second messenger¹ has stimulated interest in the chemistry of inositol phosphates. Initially, most effort was put into biological aspects of this discovery² and into the synthesis of natural inositol phosphates.³ Recently, the focus of synthetic activity has been drifting towards structurally modified *myo*-inositol phosphates with novel biological properties.⁴ Ins(1,2,6)P₃ is an inositol trisphosphate regioisomer,⁵ produced in kg quantities (Perstorp Pharma, Sweden) by partial degradation of phytic acid with phytase.⁶ Ins(1,2,6)P₃ exhibits biological activity such as inhibition of inflammatory reactions and edema in skin burn injury,⁷ and is also effective in treating acute abnormalities of nerve function in early experimental diabetes.⁸

This work describes the synthesis of the modified derivative of Ins(1,2,6)P₃ containing three carboxymethylphosphonate groups, -O-P(O)(OH)CH₂COOH. Several examples of phosphonate analogues of *myo*-inositol phosphates have been described: 5-methylenephosphonate analogue of Ins(1,4,5)P₃,⁹ 5-methylphosphonate and 5-(difluoromethyl)phosphonate analogues of Ins(1,4,5)P₃ and Ins(1,3,4,5)P₄,¹⁰ rac. 3-methylphosphonate analogues of Ins(3,4)P₂ and Ins(1,3,4)P₃,¹¹ methylphosphonate analogue of D-Ins(1)P,¹² rac. *myo*-inositol-1-O-methylphosphonate-4,5-bisphosphate,¹³ rac. *myo*-inositol 1,4,5-tris(methylphosphonate), *myo*-inositol 4,5-bis(methylphosphonate) and *myo*-inositol 5(methylphosphonate).¹⁴ Synthetic procedures leading to this type of modified phosphoinositols involve either bis[6(alkyl)benzotriazole-1-yl] alkylphosphonates¹⁰⁻¹³ or triethylammonium hydrogen methylphosphinate.¹⁴

Our main target compound was *myo*-inositol 1,2,6-tris(carboxymethylphosphonate), hexasodium salt, **11**. Other targets were *myo*-inositol-1,2-bis(carboxymethylphosphonate), tetrasodium salt, **10** and α -methyl mannopyranoside-

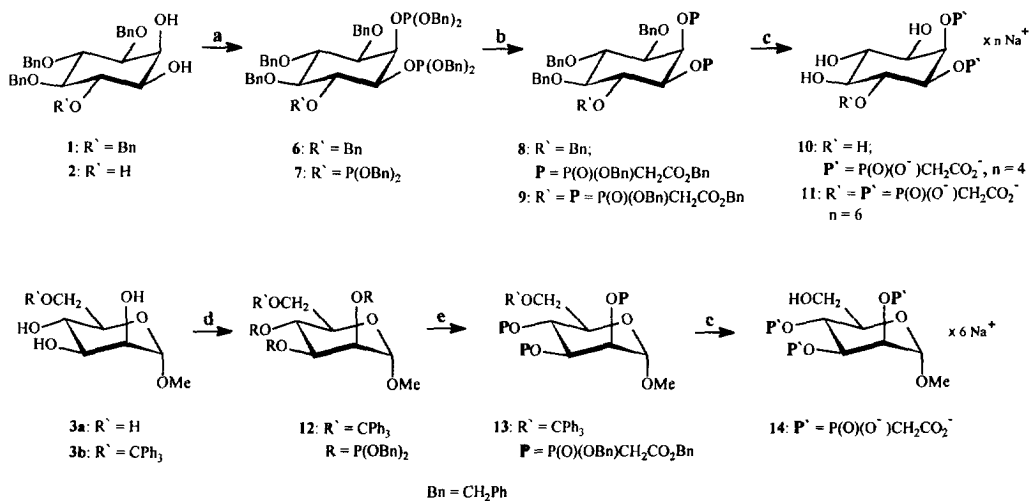
2,3,4-tris(carboxymethylphosphonate) hexasodium salt, **14**. The starting materials for the syntheses were: commercial α -methyl mannopyranoside **3a**, the relatively readily available 3,4,5,6-tetra-O-benzyl *myo*-inositol **1**¹⁵ and 3,4,5-tri-O-benzyl *myo*-inositol **2**.¹⁶

For the synthesis of the target compounds we chose a new strategy involving the following reaction sequence:

- (i) Phosphitylation of the *myo*-inositols (sugar), duly protected by benzyl groups to form inositol (sugar) dibenzyl phosphites;
- (ii) Arbuzov reaction of the *myo*-inositol (sugar) phosphites with benzyl bromoacetate;
- (iii) Deprotection of all benzyl groups.

The starting materials **1**, **2** and **3b** were phosphitylated by known phosphitylating reagents, **17** $R_2N-P(OBn)_2$ **4**: $R=Et$ or **5**: $R=Pr$.¹⁷ Yields and rates of the phosphitylation by both reagents are comparable. Phosphites **6**, **7** and **12** were purified by a short column silica-gel chromatography and characterized by ³¹P NMR.¹⁸

Arbuzov reaction of **6**, **7** and **12** with benzyl bromoacetate was carried out at 75°C - 80°C. An excess of benzyl bromoacetate acted as solvent (Scheme 1).



Scheme 1. Reagents and conditions: a) **4** or **5**, (3 eq. for **1**, 4.6 eq. for **2**), CH_2Cl_2 (2-5 ml/1 mmol of **1** or **2**), tetrazole (5 eq. for **1**, 6 eq. for **2**), r.t., 2h, 71-90%; b) $BrCH_2COOBn$ (excess), 80°C, 1-2h, 71-85%; c) Pd/C, H_2 , methanol, 12h, r.t. then NaOH aq., 71-92%; d) **4**, (4.5 eq.), CH_2Cl_2 (4 ml/1 mol of **3b**), tetrazole (6 eq.), r.t. 1h, 72%; e) $BrCH_2COOBn$ (excess), 75°C, 12h, 45%.

The reaction products **8**, **9** and **13** were purified by repeated column chromatography (Silica-gel, $CHCl_3$ - CH_2Cl_2 -acetone, 15:15:1).

The structure of **8**, **9** and **13** were confirmed by ³¹P NMR spectroscopy.¹⁹

^{31}P , ^{13}C and ^1H NMR spectra of **10**, **11** and **14** are listed in reference (20).

In the view of the mentioned above biological activity of α -trinositol^{7,8} it seemed useful to test this property of the compound **11** in the similar, following assays:²¹

- (i) Irwin test,²²
- (ii) inhibition of edema,²³
- (iii) acetic acid-induced writhing,²⁴
- (iv) shock sensitivity.²⁵

Myo-inositol 1,2,6-tris(carboxymethylphosphonate) hexasodium salt **11** and the parent α -trinositol exhibited no change in the Irwin test (mice) at dose 256 mg/kg; At dose 512 mg/kg both compounds resulted in the death of one in three mice. In the test of inhibition of edema both compounds showed at dose 64 mg/kg a statistically significant effect (95%). For phosphonate **11** no statistically significant effect was seen in acetic acid-induced writhing whereas α -trinositol exhibited an activity in this test. Neither phosphonate **11** nor α -trinositol were active in the shock sensitivity test.

References and notes

1. Berridge, M.J.; Irvine, R.F. *Nature* **1984**, *312*, 315-321; Berridge, M.J.; Irvine, R.F. *Nature* **1989**, *341*, 197-205.
2. Shears, S.B. *Biochem.J.* **1989**, *260*, 313-324; Shears, S.B. in *Advances in Second Messenger and Phosphoprotein Research* (Ed.: J.W.Putney, Jr.), Raven, New York **1992**, pp. 63-92.
3. Billington, D.C. *The Inositol Phosphates: Chemical Synthesis and Biological Significance*, VCH, Weinheim, **1993**.
4. Potter, B.V.L.; Lampe, D. *Angew.Chem. Int.Ed.Engl.* **1995**, *34*, 1933-1972.
5. Chaudhary, A.; Dorman, G.; Perstwich, G.D. *Tetrahedron Lett.* **1994**, *35*, 7521-7524.
6. Siren, M. *EPO* 179440 (1990); Blum, C.; Karlsson, S.; Schewer, G.; Spiess, B.; Rehnberg, N. *Tetrahedron Lett.*, **1995**, *36*, 7239-7242.
7. Siren, M.; Linne, L.; Persson, L. In „Inositol Phosphates and Derivatives - Synthesis, Biochemistry and Therapeutic Potential”; A.B.Reitz, Ed.; ACS Series: Washington, DC, **1991**, vol. 463, pp. 103-110.
8. Carrington, A.L.; Calcutt, N.A.; Ettlenger, C.B.; Gustafsson, T.; Tomlinson, D.R.; *Eur.J.Pharmacol.* **1993**, *237*,
9. Falck, J.R.; Abdali, A.; Wittenberger, S.J. *J.Chem.Soc., Chem.Commun.* **1990**, 953-954.
10. a) Dreef, C.E.; Schiebler, W.; van der Marel, G.A.; van Boom, J.H. *Tetrahedron Lett.* **1991**, *32*, 6021-6024; b) Dreef, C.E.; Jansze, J.P.; Elie, C.J.J.; van der Marel, G.A.; van Boom, J.H. *Carbohydr.Res.* **1992**, *234*, 37-50.
11. Dreef, C.E.; Tuinman, R.J.; Lefeber, A.W.M.; Elie, C.J.J.; van der Marel, G.A.; van Boom, J.H. *Tetrahedron* **1991**, *47*, 4709-4722.
12. Dreef, C.E.; Douwes, M.; Elie, C.J.J.; van der Marel, G.A.; van Boom, J.H. *Synthesis*, **19** 443-447.
13. Schmitt, L.; Spiess, B.; Schlewer, G. *Tetrahedron Lett.* **1993**, *34*, 7059-7060.
14. Willems, H.A.M.; Veeneman, G.H.; Westerduin, P. *Tetrahedron Lett.* **1992**, *33*, 2075-2078; Westerduin, P.; Willems, H.A.H.; van Boeckel, C.A.A. *Carbohydr.Res.* **1992**, *234*, 131-140.

15. Gigg, R.; Warren, C.D. *J.Chem.Soc. (C)*, **1969**, 2367-2371;
16. Desai, T.; Fernandez-Mayoralas, A.; Gigg, J.; Gigg, R.; Payne, S. *Carbohydr.Res.* **1990**, *205*, 105-123. The authors thank Dr. R.Gigg (National Institute for Medical Research, Mill Hill, London NW7 1AA) for generous providing them with the samples of **1** and **2**.
17. Yu.K.-L.; Fraser-Reid, B. *Tetrahedron Lett.* **1988**, *29*, 979-982; Perich, J.W.; Johns, R.B. *Tetrahedron Lett.* **1987**, *28*, 101; Uhlmann, E.; Engels, J. *Tetrahedron Lett.* **1986**, *27*, 1023; Tanaka, T.; Tamatsukuri, S.; Ikehara, T. *Tetrahedron Lett.* **1986**, *27*, 199.
18. **6** δ_p (C_6D_6): 140.4 (J_{AB} 1.8 Hz, 2P);
7 δ_p (C_6D_6): 140.2 (d, $^5J_{P1P2}$ 2.1 Hz, P-2); 140.8 (dd, $^5J_{P1P2}$ 2.1 Hz, $^5J_{P1P6}$ 3.9 Hz, P-1); 143.2 (d, $^5J_{P1P6}$ 3.9 Hz; P-6).
12 δ_p (C_6D_6): 140.3; 141.6 (2P).
19. **8** ^{31}P NMR δ_p (C_6D_6): 20.8-22.6 ppm (m).
9 ^{31}P NMR δ_p (C_6D_6): 21.0 - 22.5 (m), 24.0, 24.3, 24.4 ppm.
13 ^{31}P NMR δ_p (C_6D_6): 20.5 - 23.5 ppm (m).
20. **12** ^{31}P NMR δ_p (C_6D_6): 19.71, 19.04 ppm.
 ^{13}C NMR δ : 178.4 (t, $^2J_{p-c}$ 5.2 Hz); 79.2 (d, $^2J_{p-c}$ 7.0 Hz); 76.8 (d, J unmeasured); 76.2 (s); 74.8(s); 73.5 (d, J_{p-c} 5.2); 73.2(s); 51.0 (CH₃OH); 41.1 (d, $^1J_{p-c}$ 123.8 Hz); 40.2 (d, $^1J_{p-c}$ 122.1 Hz).
 1H NMR δ : 4.67 (dt, 1H, H-2); 4.01 (dddd, partially overlaped, 1H, H-1); 3.89 (t, 1H, H-6); 3.75 (t, 1H, H-4); 3.50 (ddd, 1H, H-3); 3.33 (t, 1H, H-5); 2.88-2.62 (m, 4H, P-CH₂). $J_{1,2}=J_{2,3}=2.5$ Hz, $J_{1,6}=J_{5,6}=J_{4,3}=9.5$ Hz; $^3J_{p-o-H2}=9.3$ Hz; $^4J_{p-o-C4H4}=1$ Hz, $^3J_{p-o-H1}=9.5$ Hz; $^2J_{p-c-H}=21.6$ Hz; $^2J_{p-c-H}=19.2$ Hz.
13 ^{31}P NMR δ (D_2O): 18.88; 19.37; 20.70 ppm.
 ^{13}C NMR δ : 178.4-178.7 (m); 78.90(s); 78.46; 78.52; 75.68; 74.40; 73.15; 42.14 (m, J_{p-c} 19 Hz); 39.78 (m).
 1H NMR δ : 4.65 (dm, $^3J_{pH}$ 10 Hz, 1H); 4.30 (dd, J 9.0 and 9.5 Hz, 1H); 4.10 (t, J 9.0 Hz, 1H); 3.78 (t, J 9.5 Hz, 1H); 3.53 (m, 2H), 3.33; 2.6-3.0 (m, 6H, P-CH₂).
14 ^{31}P NMR δ (D_2O): 21.29; 20.68; 19.69 ppm.
21. The studies were done at ITEM-labo, Le Genest-St.-Isle, France.
22. Irwin, S. *Psychopharmacologica*, **1968**, *13*, 222-257. Irwin test is a systematic observational method, used for comprehensively assessing and quantifying the behavioral and physiologic state of mouse and its response to drugs.
23. Winter, C.A.; Risley, E.A.; Nuss, G.W. *Proc.Soc.Ex Biol.Med.* **1962**, *111*, 544-547. This test is used for screening of antiinflammatory drugs.
24. Collier, H.O.J.; Dinneen, L.C.; Johnson, L.A.; Schneider, C. *Br.J.Pharmac.Chemother.* **1968**, *32*, 295-310. The test allows to evaluate an activity of drugs as inhibitors of the abominal constriction response, induced by noxious agents, in this case acetic acid.
25. Charpentier, J.C.R. *Soc.Biol.* **1961**, *155*, 727. The method is used to evaluate the analgesic effects of drugs.

(Received in UK 16 September 1996; revised 2 December 1996; accepted 6 December 1996)